



## How to submit novel alleles to Genbank and IMGT For use with AlloSeq Assign v1.0

TEC745

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## 1. Purpose

- 1.1. To submit novel alleles to IMGT they must first be submitted to Genbank, ENA or DDBJ.
- 1.2. This document describes the steps for generating sequences and feature locations for submission to Genbank.

## 2. Selecting a good allele candidate for submission

- 2.1. The allele is a good candidate if it is completely phased and homozygous.

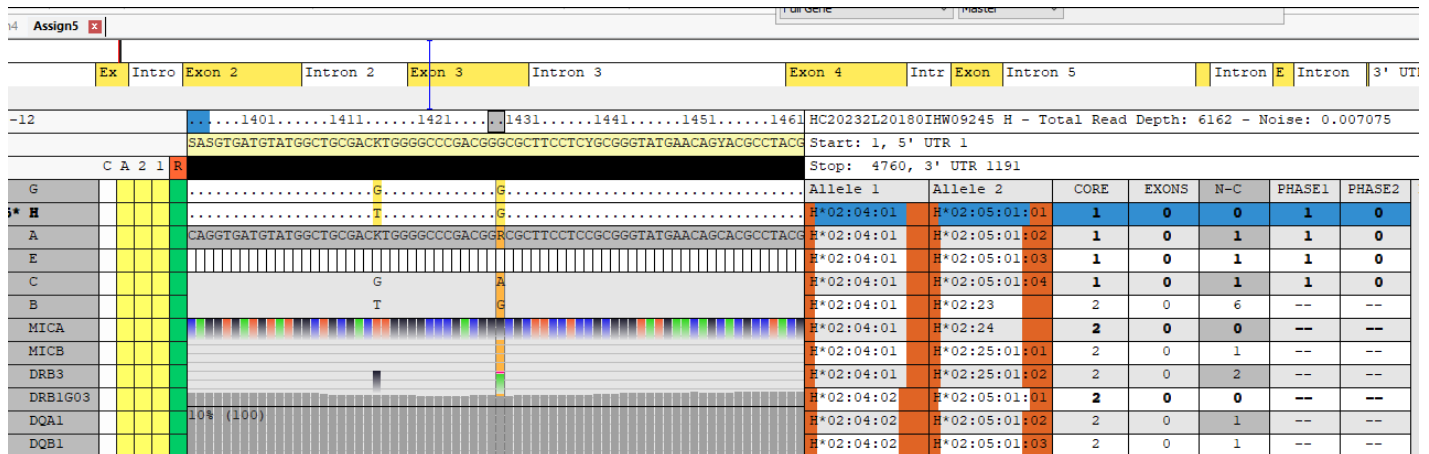


Figure 1. An example of a novel HLA-H allele with fully phased sequence.

- 2.2. If the sample is partially phased, a good allele candidate should have identical sequence for both alleles in the region that is not phased.

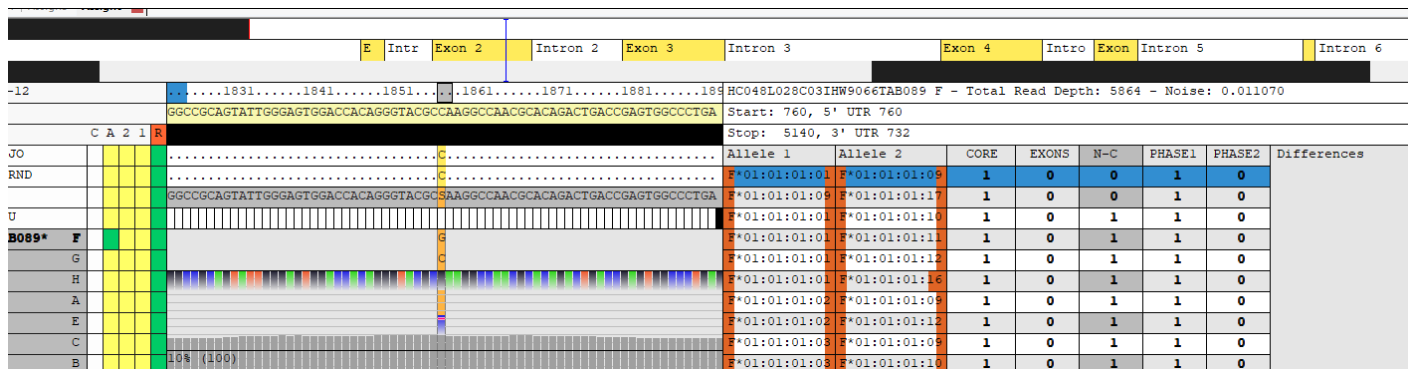


Figure 2. An example of a novel HLA-F allele with partially phased sequence. The sequence for exons 4-7 is homozygous.

**2.3. A good allele candidate may also have complete coverage with a single heterozygous position.**



Figure 3. An example of a novel HLA-E sample with a single heterozygous position.

### 3. Check the sequence for errors

### 3.1. Check for any base call errors and make edits as required.

### 3.2. Edit any masked positions.

**3.2.1.** Masked positions are excluded from analysis will report as “N” in the fasta sequence report.

**3.2.2.** Editing these positions where a clear base call can be made is recommended.

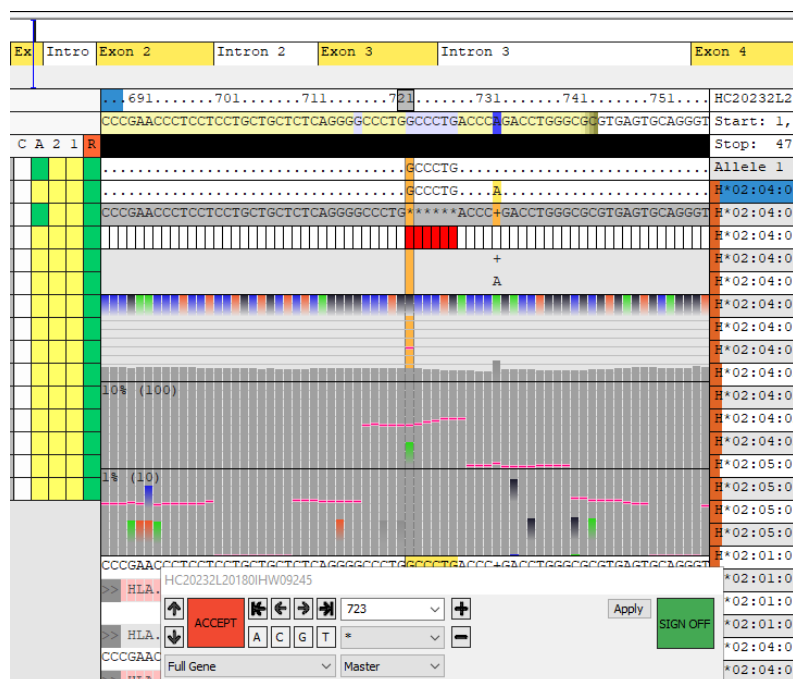


Figure 4. An example of a masked position. Masked positions are indicated by stars (\*) in the sample consensus sequence.

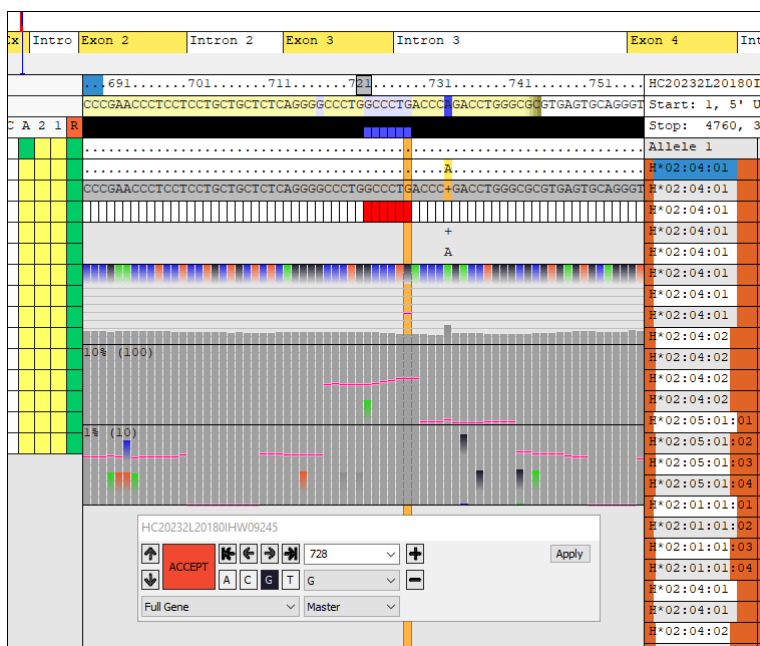


Figure 5. The sequence after editing the masked base calls.

## 4. Get the fasta sequence

Once you are satisfied with the accuracy of your sequence, use the fasta report function from Assign to generate the fasta sequence for submission.

### 4.1. To generate the fasta sequence from Assign:

**4.1.1.** Open the reports menu and select the FASTA tab.

**4.1.2.** Select the sample name from the drop down

**4.1.3.** Select the group from the dropdown. Choose **\_All\_** to generate the sequence for the full gene, or cDNA to generate the sequence for exons only.

**4.1.4.** Click generate report.

**4.1.5.** Enter the name for the file and chose the location to save the file.

NOTE 1: Selecting the locus from the locus dropdown may generate a blank report for some genes. It is recommended to select **\_All\_** in the locus dropdown and copy the sequence for the desired locus to a new file.

NOTE 2: When generating the sequence for the full gene, the start and end of the sequence will be automatically padded with "N's" due to the masks in the 5' and 3' UTRs. These will need to be manually removed before locating the gene features.

Genbank will not accept non-IUPAC codes.

NOTE 3: It is recommended to submit only cDNA sequence for class II MICA and MICB due to limited non-coding sequence available with the AlloSeq Tx 17.1 assay.

### 4.2. Fasta sequence output from AlloSeq Assign

**4.2.1.** The fasta file output from Assign should be opened with a text editor such as Notepad.

**4.2.2.** The fasta file will contain sequences for each sample and gene as selected in standard fasta format, which consists of the header

>SampleIdentifier\_IMGT/gene\_Phase #

And the sequence starting on the following line.

NOTE: Update the header as desired. A suggested format for the novel sequence header is

>SampleID gene\*##.##:new

Where ##.##:new is the local name for the novel allele.

**4.2.3.** A single sequence will be generated for homozygous loci.

**4.2.4.** For heterozygous samples 3 sequences will be generated for 1 locus:

1. Master- the sample consensus sequence with insertions expanded.
2. Phase 1- the sequence for phase layer 1, with insertions indicated by "+", homozygous deletions indicated as "-" and heterozygous deletions indicated in lowercase. This may correspond to allele 1 or 2, depending on how the phase track has been built.
3. Phase 2- the sequence for phase layer 2, with insertions indicated by "+", homozygous deletions indicated as "-" and heterozygous deletions indicated in lowercase. This may correspond to allele 1 or 2, depending on how the phase track has been built.

## 5. Indels in fasta sequence outputs from Assign

5.1. Due to how indels are handled in Assign, in certain cases it is necessary to edit indel locations in the fasta sequence outputs.

**5.1.1.** Figure 6 shows an example of a homozygous deletion in the Assign coverage pane.



Figure 6. Sample with homozygous deletion in Assign.

**5.1.2.** Figure 7 shows the fasta sequence output for the same sample. Note that the master layer sequence does not include the deletion, but phase 1 and phase 2 indicate the deletion as three dashes (---). These dashes will need to be deleted from the fasta sequence before submission.

```

>HC20223L20174C02163UCLA1000_IMG1/DQA1
ATGATCCTAAACAAGCTCTGCTGCTGGGGCCCTTGCCCTGACCACCGTGATGAGCCCTGTGGAGGTGAAGACATTGTGGCTGACCAYGTTGCCTCTT
AYGGTGTAACCTTGACCACTCTACGGTCCCTCTGGCCAGTTTACCCATGAATTTGATGGAGACGAGSAGTTCTAYGTGGACCTGGRGARGAAGGAGAC
TGCTGGWRKTTGCCTSTKYTCMRMMRAYTTAGATTTACCCGCAATTTGCACTGACAAACATCGCTGTGMYAAAAACAYAACCTGAACATCCTGATTAAA
CGCTCCAACCTTACCGCTGCTACCAATGAGGTTCTGAGGTCAACAGTGTTCCTCAAGTCTCCCGTGACRCTGGGTGACCCCAACACCTCATCTGCTTG
TGGACAACATCTTTCTCTGCTGGTCAACATCAGTGGCTGAGCAATGGGCACTCAGTCACAGAAGGTGTTTCTGAGACCAGCTTCTCTCCAAGAGTGA
TCATTCCTCTTCAAGATCAGTTACCTCACCTTCTCTCTCTGCTGATGAGATTATGACTGCAAGGTGGAGCACTGGGGCTGGAYGAGCCTCTCTG
AAACACTGGGAGCCTGAGATTCAGCMCCATGTCAGAGCTCACAGAGACTGTGGTCTGYGCCCTGGGRTTGTCTGTGGGCTCTGTGGGCAATTGTGGTGG
GSACYGTCTTSATCATCCGAGGCTGCGTTTCACTTGGTGCTTCCAGACACCAAGGGCCCTTGTGA
>HC20223L20174C02163UCLA1000_IMG1/DQA1_Phase 1
ATGATCCTAAACAAGCTCTGCTGCTGGGGCCCTTGCCCTGACCACCGTGATGAGCCCTGTGGAGGTGAAGACATTGTGGCTGACCACGTTGCCTCTT
ACGGTGTAACCTTGACCACTCTACGGTCCCTCTGGCCAGTTTACCCATGAATTTGATGGAGACGAGGAGTTCTATGTGGACCTGGAGAGGAAGGAGAC
TGCTGGAAGTTGCCTCTGTTCCAGACTTGA---TTTACCCGCAATTTGCACTGACAAACATCGCTGTGCTAAACATAACTTGAACATCCTGATT
AAACGCTCCAACCTTACCGCTGCTACCAATGAGGTTCTGAGGTCAACAGTGTTCCTCAAGTCTCCCGTGACACTGGGTGACCCCAACACCTCATCTGCT
TTGTGGACAACATCTTTCTCTGCTGGTCAACATCAGTGGCTGAGCAATGGGCACTCAGTCACAGAAGGTGTTTCTGAGACCAGCTTCTCTCCAAGAG
TGATCATTCTCTTCAAGATCAGTTACCTCACCTTCTCTCTCTGCTGATGAGATTATGACTGCAAGGTGGAGCACTGGGGCTGGATGAGCCTCTT
CTGAAACACTGGGAGCCTGAGATTCAGCACCTATGTCAGAGCTCACAGAGACTGTGGTCTGTGCCCTGGGTTGTCTGTGGGCTCTGTGGGCAATTGTGG
TGGGACCGCTTGTATCATCCGAGGCTGCGTTTCACTTGGTGCTTCCAGACACCAAGGGCCCTTGTGA
>HC20223L20174C02163UCLA1000_IMG1/DQA1_Phase 2
ATGATCCTAAACAAGCTCTGCTGCTGGGGCCCTTGCCCTGACCACCGTGATGAGCCCTGTGGAGGTGAAGACATTGTGGCTGACCATGTTGCCTCTT
ATGGTGTAACCTTGACCACTCTACGGTCCCTCTGGCCAGTTTACCCATGAATTTGATGGAGACGAGCAGTTCTACGTGGACCTGGGGAAGAAGGAGAC
TGCTGGTGTGCTGTTCTCAGACAATTTGA---TTTACCCGCAATTTGCACTGACAAACATCGCTGTGACAAAACACAACCTTGAACATCCTGATT
AAACGCTCCAACCTTACCGCTGCTACCAATGAGGTTCTGAGGTCAACAGTGTTCCTCAAGTCTCCCGTGACGTGGGTGACCCCAACACCTCATCTGCT
TTGTGGACAACATCTTTCTCTGCTGGTCAACATCAGTGGCTGAGCAATGGGCACTCAGTCACAGAAGGTGTTTCTGAGACCAGCTTCTCTCCAAGAG
TGATCATTCTCTTCAAGATCAGTTACCTCACCTTCTCTCTCTGCTGATGAGATTATGACTGCAAGGTGGAGCACTGGGGCTGGACGAGCCTCTT
CTGAAACACTGGGAGCCTGAGATTCAGCCCCATGTCAGAGCTCACAGAGACTGTGGTCTGCGCCCTGGGATTGTCTGTGGGCTCTGTGGGCAATTGTGG
TGGGCACTGTCTTATCATCCGAGGCTGCGTTTCACTTGGTGCTTCCAGACACCAAGGGCCCTTGTGA

```

Figure 7. Fasta output for Assign indicating the locations of the deletions in the 3 layers.

**5.1.3.** Figure 8 shows an example of a homozygous insertion in the Assign coverage pane.

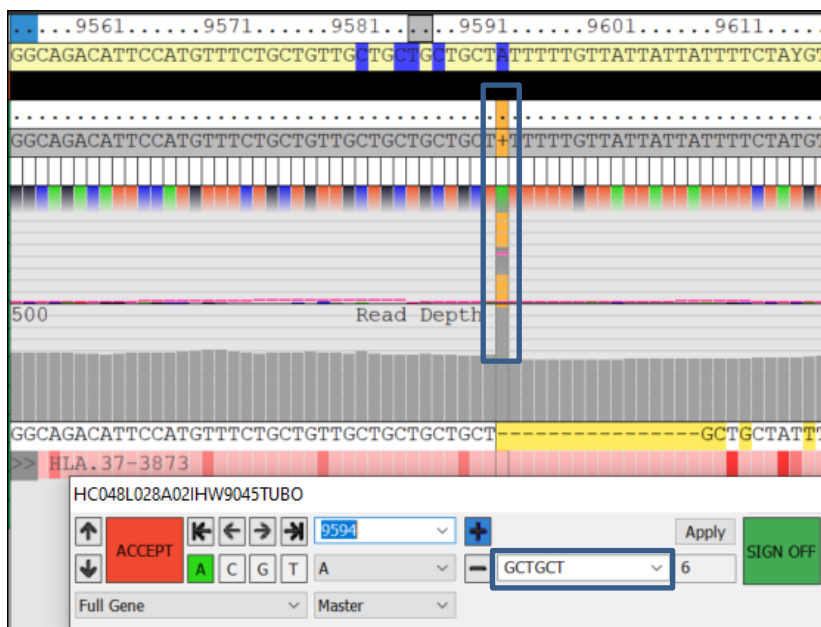


Figure 8. Sample with homozygous insertion in Assign.

**5.1.4.** Figure 9 shows the fasta sequence output for the same sample. As the sample is homozygous for the gene, a single fasta sequence is output, and the inserted bases are expanded. No edits are required for this sequence.

















## 6. Locate the gene features

Gene features include the locations of the start and end of the exons in the context of the fasta sequence.

### 6.1. cDNA sequence

**6.1.1.** In IMGT, align the closest matched allele to the novel. <https://www.ebi.ac.uk/ipd/imgt/hla/align.html>

**6.1.2.** For best results, use the closest allele as the reference sequence and align a single allele. Align in blocks of 10 bases.

**6.1.3.** follow the steps in the alignment tool.

#### STEP 1 - Select the locus and features to align

Locus:

MICA

Features:

Nucleotide - CDS

#### STEP 2 - Specify reference and required sequences

Reference sequence:

009:01:07

Specific sequences required (separated by a new line or a comma):

009:01:06

Figure 18. Alignment form in IMGT.



cDNA	10	20	30	40	50	60	70	80	90	100
MICA*009:01:07	ATGGGGCTGG	GCCCCGCTTT	CCTGCTTCTG	GCTGGCACTC	TCCCTTTTTC	ACCTCCGSGA	GCTGCTGCTG	AGCCCCACAG	TCTTCGTTAT	AACCTCACGG
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	110	120	130	140	150	160	170	180	190	200
MICA*009:01:07	TGCTGTCTGG	GGATGGATCT	GTGCAGTCAG	GGTTTCTTGC	TGAGGTATAT	CTGGATGGTC	AGCCCTTCCT	GCGCTATGAC	AGGCAGAAAT	GCAGGGCAAA
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	210	220	230	240	250	260	270	280	290	300
MICA*009:01:07	GCCCCAGGGA	CAGTGGGCGA	AAGATGTCTC	GGGAAATAAG	ACATGGGACA	GAGAGACCAG	GGACTTGACA	GGGAACGGAA	AGGAACCTCAG	GATGACCTTG
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	310	320	330	340	350	360	370	380	390	400
MICA*009:01:07	GCTCATATCA	AGGACCAGAA	AGAAG GCTTG	CATTCCCTCC	AGGAGATTAG	GGTCTGTGAG	ATCCATGAAG	ACAACAGCAC	CAGGAGCTCC	CAGCATTTCT
MICA*009:01:06	-----	-----	----- -----	-----	-----	-----	-----	-----	-----	-----
cDNA	410	420	430	440	450	460	470	480	490	500
MICA*009:01:07	ACTACGATGG	GGAGCTCTTC	CTCTCCCAA	ACGTGGAGAC	TGAGGAATGG	ACAGTGCCCC	AGTCCTCCAG	AGCTCAGACC	TTGGCCATGA	ACGTGAGGAA
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	510	520	530	540	550	560	570	580	590	600
MICA*009:01:07	TTTCTTGAGG	GAGATGCCCA	TGAAGACCAA	GACACACTAT	CACGCTATGC	ATGCAGACTG	CCTGCAGGAA	CTACGGCGAT	ATCTAGAATC	CAGCGTAGTC
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	610	620	630	640	650	660	670	680	690	700
MICA*009:01:07	CTGAGGAGAA	CAG TGCCCCC	CATGGTGAAT	GTACCCCGCA	GCGAGGCCTC	AGAGGGCAAC	ATCACCGTGA	CATGCAGGGC	TTCCAGCTTC	TATCCCCGGA
MICA*009:01:06	-----	--- -----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	710	720	730	740	750	760	770	780	790	800
MICA*009:01:07	ATATCACACT	GACCTGGCGT	CAGGATGGGG	TATCTTTGAG	CCACGACACC	CAGCAGTGGG	GGGATGTCTC	GCTGATGGG	AATGGAACCT	ACCAGACCTG
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	810	820	830	840	850	860	870	880	890	900
MICA*009:01:07	GGTGGCCACC	AGGATTGGCC	AAGGAGAGGA	GCAGAGGTTT	ACCTGCTACA	TGGAACACAG	CGGGAATCAC	AGCACTCACC	CTGTGCCCTC	TG GGAAAGTG
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-- -----
cDNA	910	920	930	940	950	960	970	980	990	1000
MICA*009:01:07	CTGGTGCTTC	AGAGTCATTG	GCAGACATTC	CATGTTTCTG	CTGTTGCTGC	TGCTGCTGCT	GCTATTTTTC	TTATTATTAT	TTTCTATGTC	CGTTGTTGTA
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	1010	1020	1030	1040	1050	1060	1070	1080	1090	1100
MICA*009:01:07	AGAAGAAAAC	ATCAGCTGCA	GAGGGTCCAG	AGCTCGTAG	CCTGCAGGTC	CTGGATCAAC	ACCCAGTTGG	GACGAGTGAC	CACAGGGATG	CCACACAGCT
MICA*009:01:06	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----
cDNA	1110	1120	1130	1140	1150	1160				
MICA*009:01:07	CGGATTTCAG	CCTCTGATGT	CAGCTCTTGG	GTCACTGGC	TCACTGAGG	GCGCCTAG				
MICA*009:01:06	-----	-----	-----	-----	-----	-----				

Figure 19. Alignment of MICA alleles in IMGT.

**6.1.4.** Use the numbering in the alignment to determine the feature locations. The vertical lines (Pipes) indicate the exon boundaries.

Table 1. Feature locations for MICA as determined from the alignment.

Feature	Location
Exon 1	1-70
Exon 2	71-325
Exon 3	326-613
Exon 4	614-892
Exon 5	893-1030
Exon 6	1031-1150

## 6.2. gDNA sequence

- 6.2.1. To determine the feature locations, BLAST the fasta sequence output from Assign using IMGT BLAST [https://www.ebi.ac.uk/Tools/services/web\\_ncbiblast/toolform.ebi?tool=ncbiblast&context=nucleotide&database=imgthlagen/](https://www.ebi.ac.uk/Tools/services/web_ncbiblast/toolform.ebi?tool=ncbiblast&context=nucleotide&database=imgthlagen/)
- 6.2.2. Select the appropriate IMGT database to search. i.e., for a full gene sequence select IMGT> IMGT/HLA (genomic).
- 6.2.3. Paste the fasta nucleotide sequence for the novel allele into IMGT Nucleotide BLAST and submit the job.

**NCBI BLAST+**

Protein | **Nucleotide** | Vectors | Web services | Help & Documentation | Bioinformatics Tools FAQ | Feedback | Share

Tools > Sequence Similarity Searching > NCBI BLAST

Results for job ncbiblast-l20210819-035416-0884-15002771-p2m

Summary Table | Tool Output | Visual Output | Functional Predictions | Result Summary | Submission Details

**Selection:**

**Apply to selection:**  
**Annotations:**

**Alignments:**

**Entries:**  
 in  
 fasta

**format**

**Tools:**  
  
 Clustal Omega

Align.	DB:ID	Source	Length	Score (Bits)	Identities %	Positives %	E()
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA00934	E*01:01:01:01 3822 bp Cross-references and related information in: Nucleotide sequences Literature	3822	7589.1	100.0	100.0	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA00938	E*01:03:01:01 3822 bp Cross-references and related information in: Nucleotide sequences Literature	3822	7581.2	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA00937	E*01:03:02:01 3822 bp Cross-references and related information in: Nucleotide sequences Literature	3822	7553.3	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA02224	E*01:03:04 3822 bp Cross-references and related information in: Nucleotide sequences Literature	3822	7545.3	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA05913	E*01:03:02:02 3822 bp Cross-references and related information in: Nucleotide sequences Literature	3822	7545.3	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA02226	E*01:03:01:02 3785 bp Cross-references and related information in: Nucleotide sequences Literature	3785	7479.9	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA02225	E*01:01:01:02 3785 bp Cross-references and related information in: Nucleotide sequences Literature	3785	7477.9	99.9	99.9	0.0
<input checked="" type="checkbox"/>	IMGTHLAgen:HLA02450	E*01:01:01:03 3738 bp Cross-references and related information in: Nucleotide sequences Literature	3738	7394.7	99.9	99.9	0.0

Figure 20. Summary table of hits for novel HLA-E sequence.

- 6.2.4. IMGT BLAST will then display the matches for the sequence.
- 6.2.5. By default, all the matches will be selected. Click Clear under Selection to deselect all matches.
- 6.2.6. Select the top result, then click show under Annotations and show under Alignments to review the results for the top result.
- 6.2.7. If the length of the Subject sequence is the same length as the novel fasta sequence (or query) then the annotations will correspond to the novel sequence. Check the length in the alignment and locate the novel position(s).



```

>IMGTHLAgene:HLA00934 E*01:01:01:01 3822 bp
Length=3822

Score = 7569 bits (3818), Expect = 0.0
Identities = 3821/3822 (99%), Gaps = 0/3822 (0%)
Strand=Plus/Plus

Query 1      CAAAGTGCTGAGATTACAGGCGTGAGCCACCGCGCCAGCCAGGACTAATTTCTAAGAGT 60
             |||
Sbjct 1      CAAAGTGCTGAGATTACAGGCGTGAGCCACCGCGCCAGCCAGGACTAATTTCTAAGAGT 60

```

Figure 21. The start of the alignment for the novel HLA-E sequence and top match E\*01:01:01:01 matches.

```

Query 3781  GGCAGAGTGCGGCAGCTCATGCCTGTAATCCCAGCACTTAGG 3822
             |||
Sbjct 3781  GGCAGAGTGCGGCAGCTCATGCCTGTAATCCCAGCACTTAGG 3822

```

Figure 22. The end of the alignment for the novel HLA-E sequence and top match E\*01:01:01:01 matches.

```

Query 2101  TGGTGCCTTCTGGAGAGGAGCAGAGATACACGTGCCATGTGCAGAAATGAGGGGCTACCCG 2160
             |||
Sbjct 2101  TGGTGCCTTCTGGAGAGGAGCAGAGATACACGTGCCATGTGCAGCAATGAGGGGCTACCCG 2160

```

Figure 23. The location of the novel SNP as indicated by the absence of the pipe in the alignment for a novel HLA-E sequence and top match E\*01:01:01:01.

**6.2.8.** The alignment will also highlight any errors in the fasta sequence due to N's or "+". If any errors are identified these can be edited in the fasta sequence.

The screenshot below shows an example of an HLA-H sequence error due to "+". The "+" has been converted to N, and there is missing sequence in the fasta file.

```

Query 3241  TTGTTCCCTGCCCTTCCCTTTGTGACTTGAAGAACCCTGA-----CTTCTTCAAAGGCACCT 3296
             |||
Sbjct 3241  TTGTTCCCTGCCCTTCCCTTTGTGACTTGAAGAACCCTGACTTTCTTCTCAAAGGCACCT 3300

```

Figure 24. Alignment of novel HLA-H\*02:04:new sequence and H\*02:04:01.

**6.2.9. Error! Reference source not found.** shows that at the position where Assign has put a “+” there is an A and 4nt insertion TTCT, matching the A\*02:04:01 allele.

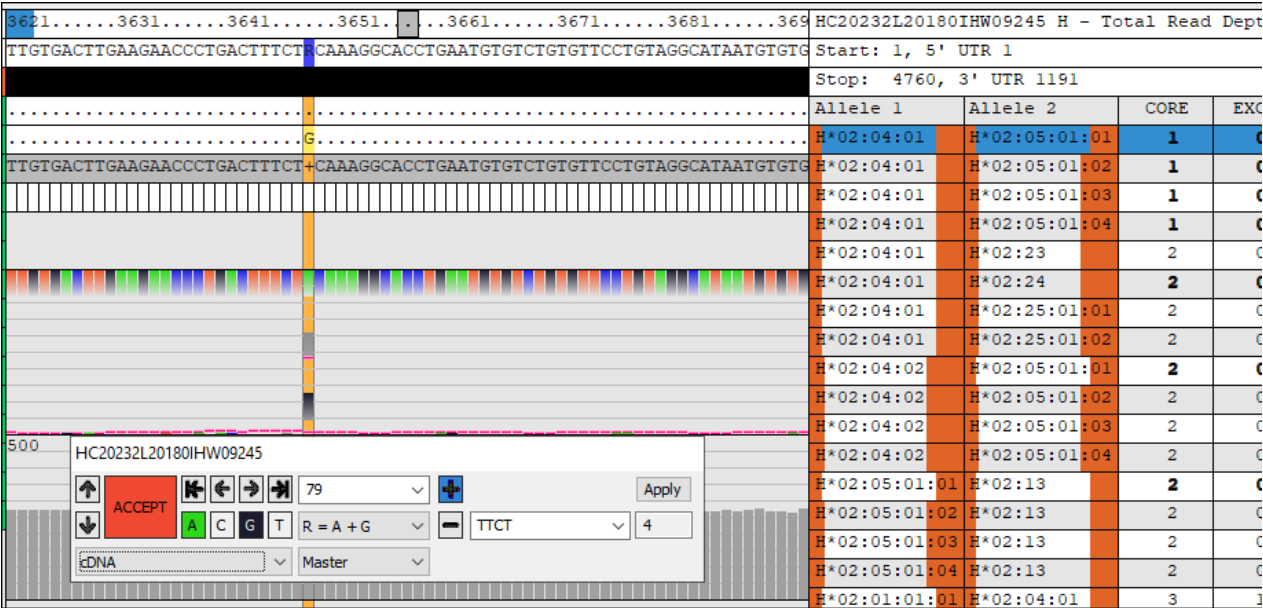


Figure 25. Alignment of a novel HLA-H sample in Assign highlighting the insertion.

**6.2.10.** This position can be edited in the fasta sequence to match the display in Assign, by locating the “+” in the fasta sequence, then replacing it with “TTCTA”.



[illegible]

Figure 27. The sequence edited to include the inserted bases in the novel HLA-H\*02:04:New allele.

- 6.2.12.** Once satisfied that the annotation locations match the novel allele sequence, they can be used to create the 5-column feature table if submitting to Genbank.

## 7. Genbank submission

### 7.1. the 5-column feature table for Genbank submission

- 7.1.1.** The Genbank Bankit tool has 2 functions- *manual entry using a form* or *upload a 5-column feature table*- for adding gene feature locations to a sequence, but the same information is required for each.
- 7.1.2.** Creating a 5-column feature table allows the inclusion of exons, CDS and details of the gene, as well as other features described in the help links below from Genbank.
- 7.1.3.** This page <https://www.ncbi.nlm.nih.gov/WebSub/html/help/feature-table.html> describes the requirements for creating the feature table.
- 7.1.4.** This page [https://www.insdc.org/files/feature\\_table.html](https://www.insdc.org/files/feature_table.html) contains detailed information on the feature table.
- 7.1.5.** 5-column feature tables are saved as .tbl or .txt format and may be edited with notepad.
- 7.1.6.** Some examples of feature table files have been included with this SOP.
- 7.1.7.** Key features of the 5-column feature table:

Example HLA-E

Example\_E01NEW.tbl - Notepad

File Edit Format View Help

>Feature Example\_E\*01:NEW

1	3822	gene	gene	HLA-E
301	364	CDS		
495	764			
1009	1284			
1906	2181			
2306	2422			
3173	3205			
3310	3350			
			product HLA-E	
			codon_start	1
301	364	exon	label	Exon 1
495	764	exon	label	Exon 2
1009	1284	exon	label	Exon 3
1906	2181	exon	label	Exon 4
2306	2422	exon	label	Exon 5
3173	3205	exon	label	Exon 6
3310	3352	exon	label	Exon 7
3518	3522	exon	label	Exon 8

The 5-column feature table as viewed in notepad.

Figure 28. 5-column feature table example for HLA-E

Example\_E01NEW.tbl - Notepad

File Edit Format View Help

>Feature Example\_E\*01:NEW

1	3822	gene	gene	HLA-E
301	364	CDS		
495	764			
1009	1284			
1906	2181			
2306	2422			
3173	3205			
3310	3350			
			product HLA-E	
			codon_start	1
301	364	exon	label	Exon 1
495	764	exon	label	Exon 2
1009	1284	exon	label	Exon 3
1906	2181	exon	label	Exon 4
2306	2422	exon	label	Exon 5
3173	3205	exon	label	Exon 6
3310	3352	exon	label	Exon 7
3518	3522	exon	label	Exon 8

Column descriptions:

1.Sequence Identifier

Line 1

2. Column 1: Start location (first nucleotide) of a feature

3. Column 2: Stop location (last nucleotide) of a feature

4. Column 3: Feature name (for example, 'CDS' or 'mRNA' or 'rRNA' or 'gene' or 'exon')

Line 2

5. Column 4: Qualifier name (for example, 'product' or 'number' or 'gene' or 'note')

6. Column 5: Qualifier value

Figure 29. 5-column feature table with columns indicated.

Example\_E01NEW.tbl - Notepad

File Edit Format View Help

>Feature Example E\*01:NEW

1	3822	gene	gene	HLA-E
301	364	CDS		
495	764			
1009	1284			
1906	2181			
2306	2422			
3173	3205			
3310	3350			
			product HLA-E	
			codon_start	1
301	364	exon	label	Exon 1
495	764	exon	label	Exon 2
1009	1284	exon	label	Exon 3
1906	2181	exon	label	Exon 4
2306	2422	exon	label	Exon 5
3173	3205	exon	label	Exon 6
3310	3352	exon	label	Exon 7
3518	3522	exon	label	Exon 8

Feature examples:

1. Gene Feature
2. CDS Feature
3. Exon Features

Figure 30. 5-column feature table with features indicated.

**7.1.8.** Tab stops are used to separate each column in the feature table.

## 7.2. Create the 5-column feature table

### 7.2.1. For cDNA sequences- feature table and gapped sequence

**7.2.2.** Use the template *TEC745-1\_Create feature table cDNA.xlsx* to create the 5-column feature table.

**7.2.3.** Enter the feature locations into the first tab- exon\_locations.

**7.2.4.** Copy the cDNA fasta sequence output for the novel allele including the header and paste into the paste\_fasta tab.

**7.2.5.** The template will add a string of N's (gaps) to the fasta sequence to represent to introns.

**7.2.6.** The updated sequence can then be copied and pasted to a new text document and saved as .fasta or .txt file.

**7.2.7.** The template will generate the 5-column feature table with updated exon locations and gap locations in the tab tbl\_output.

**7.2.8.** The feature table can then be copied and pasted into a new text document and saved as .tbl or .txt file.

### 7.2.9. For gDNA sequences- feature table

**7.2.10.** Use the template *TEC745-2\_Create feature table gDNA.xlsx* to create the 5-column feature table.

**7.2.11.** In the first tab exon\_locations, select the gene name from the list to populate the default exon locations, or manually enter the exon locations in the table.

NOTE: the default exon locations have been calculated based on common alleles and will not be suitable for all sequences. It is recommended to carefully check the feature locations for each novel sequence.



**7.2.12.** Copy the gDNA fasta sequence output including the header and paste into the paste\_fasta tab.

NOTE: the fasta sequence is used for creating the feature table only and is not modified.

**7.2.13.** The template will generate the 5-column feature table in the tbl\_output tab.

**7.2.14.** The feature table can then be copied and pasted into a new text document and saved as .tbl or .txt file.

### 7.3. NCBI account

**7.3.1.** Go to <https://www.ncbi.nlm.nih.gov/> and follow the instructions to set up an account.

### 7.4. Submit with Bankit

**7.4.1.** Click submit on the NCBI homepage and follow the prompts for submission or go directly to Bankit <https://submit.ncbi.nlm.nih.gov/about/bankit/>

**7.4.2.** Follow the prompts on the Bankit forms to enter contact information and references.

**7.4.3.** In the Sequencing Technology form, select “Other” for Method, and enter “AlloSeq Tx17.1 Hybrid Capture”. Select assembled sequences and enter “AlloSeq Assign” for the Assembly program and enter the version of AlloSeq Assign used.

**7.4.4.** In the Nucleotide form, enter the desired release date. Select genomic DNA for the molecule type, Linear topology and No to complete sequence submission. Either paste the fasta sequence for the novel allele or upload the file.

**7.4.5.** Enter the Organism name (homo sapiens), Submission category and Source modifiers as appropriate.

**7.4.6.** In the Features form, select Add features by uploading 5 column feature table. Choose the file, then upload. Once uploaded the features will populate.

**7.4.7.** Review the Submission and Submit.

## 8. IMGT submission

### 8.1. To submit to IMGT the following are required:

#### **8.1.1.** Accession number for the sequence.

8.1.1.1. This will be provided via email by Genbank once they have performed initial processing of the submitted sequence.

#### **8.1.2.** Fasta sequence

8.1.2.1. use the sequence prepared for the Genbank/ENA/DDBJ submission.

#### **8.1.3.** The name of the closest known allele

8.1.3.1. The top result as determined by the IMGT BLAST search.

#### **8.1.4.** The locations of the differences to the closest known allele.

8.1.4.1. The location of the differences to the closest known allele can be determined using the IMGT BLAST alignment, or by performing a sequence alignment of the closest known allele <https://www.ebi.ac.uk/ipd/imgt/hla/align.html>

Summary Table Tool Output Visual Output Functional Predictions Result Summary Submission Details									
<b>Selection:</b> Select All Invert Clear		Align.	DB:ID	Source	Length	Score (Bits)	Identities %	Positives %	E()
<b>Apply to selection:</b> Annotations: Show Hide		<input checked="" type="checkbox"/>	IMGTHLA:HLA02552	H*02:04:01 3510 bp Cross-references and related information in: ▶ Nucleotide sequences ▶ Literature	3510	6905.0	99.8	99.8	0.0

Figure 31. Top result for H\*02:04:NEW from the IMGT BLAST search.

Query	1021	CACACCATGCAGGTGATGTATGGCTGCGACGTGGGGCCCGACGGACCTTCCTCCGCGGG	1088
Sbjct	1021	CACACCATGCAGGTGATGTATGGCTGCGACGTGGGGCCCGACGGACCTTCCTCCGCGGG	1088

Figure 32. Alignment from the HLA-H\*02:04:NEW BLAST search indicating the novel position at nt 1065.

#### 8.1.5. The genotypes of other HLA loci such as A, B and DRB1.

8.2. Use the submission tool on the IMGT website to submit the novel allele.

8.2.1. <https://www.ebi.ac.uk/ipd/imgt/hla/subs/submit.html>

8.2.2. Once the novel sequence has been accepted and processed, IMGT will email the official name for the novel sequence.



## 9. References

### NCBI

National Center for Biotechnology Information (NCBI)[Internet]. Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information; [1988] – [cited 2022 Jan 04]. Available from: <https://www.ncbi.nlm.nih.gov/>

### IMGT

Robinson, J, Barker, DJ, Georgiou, X, Cooper, MA, Flicek, P, Marsh, SGE, The IPD-IMGT/HLA Database, Nucleic Acids Research (2020) 43:D948-D955

## 10. Resources

### DDBJ Nucleotide Sequence Submission System (NSSS)

[https://www.ddbj.nig.ac.jp/assets/files/pdf/ddbj/websubHelp\\_full-e.pdf](https://www.ddbj.nig.ac.jp/assets/files/pdf/ddbj/websubHelp_full-e.pdf)

### Submissions to GenBank

<https://www.ncbi.nlm.nih.gov/genbank/submit/>

### General Guide on ENA Data Submission

<https://ena-docs.readthedocs.io/en/latest/submit/general-guide.html#>

### AlloSeq Assign Instructions for Use

<https://labproducts.caredx.com/software/assign/alloseq-assign/manuals/>

## 11. Customer Support

Website: <https://labproducts.caredx.com/>

For Technical Support please email: [techsupport-global@caredx.com](mailto:techsupport-global@caredx.com)

For ordering details, please refer to the CareDx website: <https://labproducts.caredx.com/>

## 12.Appendix

Table 2. Amino acids

Amino Acid	DNA codons	Amino Acid	DNA codons
<b>Ala A</b>	GCT, GCC, GCA, GCG	<b>Lys K</b>	AAG, AAA
<b>Arg R</b>	CGT, CGC, CGA, CGG, AGA, AGG	<b>Met M</b>	ATG
<b>Asn N</b>	AAT, AAC	<b>Phe F</b>	TTT, TTC
<b>Asp D</b>	GAT, GAC	<b>Pro P</b>	CCT, CCC, CCA, CCG
<b>Cys C</b>	TGT, TGC	<b>Ser S</b>	TCT, TCC, TCA, TCG, AGT, AGC
<b>Gln Q</b>	CAA, CAG	<b>Thr T</b>	ACT, ACC, ACA, ACG
<b>Glu E</b>	GAA, GAG	<b>Trp W</b>	TGG
<b>Gly G</b>	GGT, GGC, GGA, GGG	<b>Tyr Y</b>	TAT, TAC
<b>His H</b>	CAT, CAC	<b>Val V</b>	GTT, GTC, GTA, GTG
<b>Ile I</b>	ATT, ATC, ATA	<b>Start</b>	ATG
<b>Leu L</b>	TTA, TTG, CTT, CTC, CTA, CTG	<b>Stop</b>	TAA, TGA, TAG

## Revision History

Version	Date	Modification
1.0	18-Aug-21	Drafted document. Issued 14 Jun 2022